

International Journal of Engineering Researches and Management Studies CONCEPT OF GLOWING PLANTS: FUTURE POSSIBILITY OF SUSTAINABLE DEVELOPMENT OF SOCIETY

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ABSTRACT

Bioluminescence is a very common physical phenomenon to generate emit light in the presence of oxygen. Bioluminescence plants represent a novel frontier in sustainable biotechnology, offering a unique convergence of natural illumination and synthetic biology. There have been transformed with bioluminescence gene to originate in living organism or plants so far – from bacteria, fungi, insects, firefly, marine organism, algae, dinoflagellate, jellyfish, earthworm, and sea star fish into plant genomes, researchers are developing self-sustaining, light-emitting plants that require no external electricity. During the past years, it was found that bioluminescence system such as the luciferase-luciferin system, PBIN technique, bacterial bioluminescence system, caffeic acid cycle (CAC), and fluorescent proteins (FPs) are generating glowing plants in the dark area and can be easily seen by our naked eyes. Here, we summarized the review of literatures rebuilding bioluminescent system to development of auto luminescence in plant cells. In future possibilities, bioluminescence plants can be used not only in scientific research area, but also a capable of alternative option as formation of artificial light source of lighting. As the next generation of society seeks innovative solutions to climate and energy challenges, bioluminescent plants offer a promising and symbolic step toward a more regenerative and ecologically integrated future.

KEYWORDS: Bioluminescent system; Genetic engineering; Sustainable development

1. INTRODUCTION

Bioluminescence reaction is most of the production of light in living organism, which was discovered among more than 10,000 species from 800 genera (Haddock *et al.*, 2010). They perform a special chemical reaction to produce emit visible light. Bioluminescence considered as a special kind of chemical signals for the living organisms. Basically, bioluminescence word is defined as the general term of a chemical compound that is biosynthesized is a luminous organism to provide energy for emission light by being oxidized by catalysis of a luciferase gene in the presence of oxygen. It is the very natural phenomenon to glow up the living organism with its secret behaviour (Shimomura 2006). Bioluminescence, the biochemical formation of light by a luciferase-luciferin chemical reaction within living organism. It is a natural phenomenon to generate by different type of proteins called luciferases that makes easy to the oxidation of substrates called luciferins. The most relatable review, focuses on the apart of scientists to perform a special role - in the region of glowing plants (Li *et al.*, 2021).

Recently; discovered some new genus of bioluminescence species in the forest of Meghalaya, India (Karunarathna *et al.*, 2020). Bioluminescence is also performed by the photoluminescent activity that glow continuously for a short or long period of time in the absence of light. The alternative strategy to develop novel bioluminescence system is emerging. Many species of fungi naturally produce light, a term known as bioluminescence or fluorescence. During the past some years, found the fluorescent gene in the protein and bioluminescence system are responsible for encourage of most native luminescence gene (Kim and Paulmurugan 2021). The presence of bioluminescent system to change the mechanism of living organisms with a physical advantage in certain ecological situations. Bioluminescence jellyfish *Ae666quorea Victoria* to expresses the green fluorescent protein (GFP) along with the photoprotein aequorin (Morise *et al.*, 1974). Bioluminescence gene was synthesized by fungal, bacteria, insects, fire fly, marine organisms, algae, dinoflagellate, jellyfish, worms, and sea star fish (John 2008).

Most of cases, light emission results from the chemical oxidation. Bioluminescence gene produce cold light in the presence of oxygen but not in higher plant on the Earth. Recently - a few plants have developed, converted using bioluminescence system such as the caffeic acid cycle, bacterial bioluminescence system, luciferase-luciferin system that allow them to glowing the darkness and can be easily seen by our naked eyes giving a clear lighting advantage, through bio-techniques, it is possible to introduce luminescence system into living plant cells as biomarkers (Mitiouchkina *et al.*, 2020). Some plants transformed with luminescence systems for example – Arugula (*Eruca sativa*), Kale (*Brassica oleracea*), Spinach (*Spinacia oleracea*), Watercress (*Nasturtium officinale*), and Tobacco (*Nicotiana tabacum*) plants using the bioluminescent system to produce glowing light (Kwak *et al.*, 2017 and Mitiouchkina *et al.*, 2020). Engineered biolumeneting plant ATP-powered



genetically encoded high autoluminescent light source as sustainable alternatives (Gordiichuk *et al.*, 2021). Might be useful not only for basic plant research but also for ornamental purpose as a novel property of glowing flower and plant cells. Development a plant with self-sustain bioluminescence gene. Autoluminescent plants engineered to express a fungal, bacterial bioluminescence system that converts caffeic acid into luciferin and report self-sustain bioluminescence that is visible to produce sufficient light. Bioluminescence results from a biochemical reaction involving a luciferase enzyme and a luciferin substrate often requiring molecular oxygen or cofactors such as ATP and NADH catalysed by the specific enzyme, luciferase (Fleiss and Sakisyan 2019). Bioluminescent plants developed through the integration of bioluminescent gene pathways from fungi, bacteria or marine organisms represent a novel and promising tool for advancing sustainable development in modern society. These genetically engineered organisms can emit visible light autonomously without requiring external energy sources making them potential alternative to conventional lighting system. Their application extends beyond aesthetic or ornamental use, offering benefits such as reduced electricity consumption, lower carbon emission, and innovative green infrastructure solutions in urban planning. Furthermore, bioluminescent plants may serve as biosensors for environmental monitoring and contributing to ecological awareness and resource management. Current advancements in synthetic biology including successful expression of fungal bioluminescent in plants (Mitiouchkina *et al.*, 2020).

Mechanism of Bioluminescence: Bioluminescence typically involves the oxidation of a substrate (luciferin), catalysed by an enzyme (luciferase), resulting in light emission. In some marine species, luciferase and luciferin are part of a single complex known as a photoprotein which requires additional cofactors to function. The colour of the emitted light depends on the specific luciferin-luciferase system and can range from blue to red (Duchatelet and Dupont 2024). Some basic principal behind the light emission is as follows -

Reaction 1: A chemical reaction produce bioluminescence activity, which is a sort autoluminescence in which light is emitted. The reaction rate for the luciferin is controlled by a luciferase enzyme and luciferin convert into oxyluciferin and generate glowing light. Luciferin is the main component in bioluminescence reaction. Luciferin from the oxidised (oxyluciferin), ATP and NADH catalysed by the specific enzyme luciferase.

Reaction 2: The bioluminescence chemical reaction in the firefly is the enzymes called luciferin and luciferase. Luciferin was converted to luciferyladenylate by ATP in the presence of Mg2+. Mg2+, ATP are required for the reaction, while adenosine monophosphate (AMP) and pyrophosphate (PP) are waste product. Luciferin

> Luciferase + D-luciferin + ATP + Mg^{2+} Luciferase-D-luciferyl-adenylate + Pyro-phosphate + Mg^{2+}

Reaction 3: Bioluminescence is due to oxygenation reactions – oxygen reacts with substances called luciferin, producing energy in the form of light. The component Luciferase-D-luciferyladenylate was oxidised by molecular oxygen and produce photons of Luciferase-oxyluciferin and light. In this process the luciferase become oxygenated to from oxyluciferin.

Luciferase-D-luciferyladenylate + O_2 \longrightarrow Luciferase-Oxyluciferin + CO_2 + AMP + Light

Reaction 4: At the core of all bioluminescent systems lies a reaction between luciferin (substrate) and luciferase (enzyme). The reaction typically involve – Oxidation of luciferin, emission of a photon (visible light), and regeneration of substrate in firefly bioluminescence operating system.

Luciferin + ATP + O_2 \longrightarrow Oxyluciferin + CO_2 + Light

Reaction 5: In some cases, additional cofactors such as FMNH2 (bacteria) are required. Aliivibrio fischeri is a marine bacterium that produces light through a luciferase-catalyzed reaction involving FMNH2, oxygen, and a long chain aldehyde. This bacterium often forms symbiotic relationship with marine animals such as the Hawaiian bobtail squid providing camouflage through counter-illumination.

$$FMNH_2 + RCHO + O_2 \longrightarrow FMN + RCOOH + H_2O + Light$$

- Luciferin : The light-emitting substrate
- **Luciferase** : The enzyme that catalyzes the reaction
- ATP : Provides energy and activates luciferin
- **Mg**²⁺ : Essential cofactor for luciferase activity
- **O2** : Required for oxidation
- **RCHO**: Long-chain aliphatic aldehyde (tetradecanal)



- **RCOOH**: Long-chain fatty acid (tetradecanoic acid or myristic acid)
- **FMN** : Oxidized flavin
- FMNH2 : Reduced flavin mononucleotide

In the present review highlights the progress report on the bioluminescent systems in plants cell and various scientist contribution to the success of glowing plants to produced emit light using the different bioluminescence system were presented.

Luminescence gene source	Engineered plants	Derived from	Gene introduces	References
Firefly	Nicotiana tabacum	Photinus pyralis	Firefly luciferase	Ow et al., (1986)
Firefly	Daucus carota	Photinus pyralis	Firefly luciferase	Ow et al., (1986)
Bacterium	Nicotiana tabacum	Photobacterium leiognathid	luxCDABEG	Krichevsky <i>et al.</i> , (2010)
Nano-lantern	Nicotinan benthamiana	Artificial nano- lantern	pH2GW7-CT nano-lantern	Saito <i>et al.</i> , (2012)
Fluorescent protein	Torenia fournieri	Chiridius poppei	CpYGFP	Sasaki <i>et al.</i> , (2014)
N. tabacum	Neonothopanus nambi	H3H, Hisps, Luz and CPH	Fungus luciferase	Mitiouchkina <i>et al.</i> , (2020)
Firefly	Brassica oleracea	Photinus pyralis	Firefly luciferase	Kwak et al., (2017)
Firefly	Spinacia oleracea	Photinus pyralis	Firefly luciferase, luciferin and CoA	Kwak et al., (2017)
Fluorescent protein	Chrysanthemum morifolium	Chiridius poppei	CpYGFP	Kishi-Kaboshi <i>et al.</i> , (2017)
Fluorescent protein	Petunia hybrida	Chiridius poppei	eYGPuv	Chin <i>et al.</i> , (2018)
Fungi	Petunia hybrida	Aspergillus nidulans and Neonothopanus nambi	NPGA, H3H, Hisps, Luz, and CPH	Khakhar <i>et al.</i> , (2020)
Fungi	Nicotiana benthamiana	Aspergillus nidulans and Neonothopanus nambi	NPGA, H3H, Hisps, Luz, and CPH	Khakhar <i>et al.</i> , (2020)
Fungi	Catharathus roseus	Aspergillus nidulans and neonothopanus nambi	NPGA, H3H, Hisps, Luz, and CPH	Khakhar <i>et al.</i> , (2020)
Rosa rubiginosa	Aspergillus nidulans and Neonothopanus nambi	NPGA, H3H, Hisps, Luz and CPH	Fungus luciferase	Khakhar <i>et al.</i> , (2020)
Nano-lantern	Arabidopsis thaliana	Artificial nano- lantern	Green enhanced nano-lantern (GeNL)	Furunata <i>et al.</i> , (2020)

Glowing plants with bioluminescent system

Bioluminescent system is very largely understudied, it's very widely use to research area. Various techniques have been developed and use to achieve this goal. Include the techniques like – microinjection, gene gun technique, liposome-mediated, electroporation technique, PBIN, spray method, and *Agrobacterium tumefaciens* method.

Up to now, five Natural bioluminescent system that origin from fireflies, bacteria, fungi, protein have been reconstructed in plants and also used by chemically reaction for glowing plants. The chemically reaction that generates emit light happens within a living organism. Plants can produce a special chemical reaction to generating light, scientists have found way to artificially replicate the phenomena.

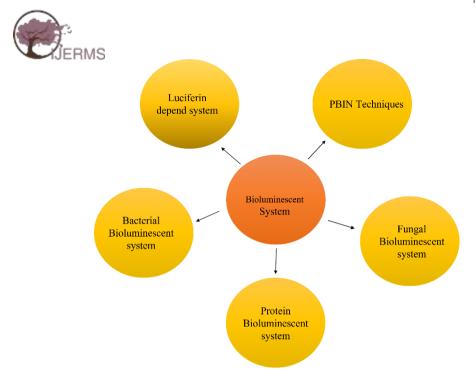


Figure 1: Five types of natural bioluminescent system

2. NATURAL BIOLUMINESCENT SYSTEM

I. D-Luciferase depend system

In fireflies, first-time bioluminescent system that was introduced the *Photinus pyralis* luciferase gene into *Daucus carota* and *N. tabacum* plants is firefly luciferase-luciferin system (Ow *et al.*, 1986). Luciferin-Luciferase system (i.e., bacterial, fungal, firefly, coelenterazine-based) have inherently different emission spectra. D-luciferase depend system is luciferase-luciferin system belong to one of the most practically important and most commonly collections of bioluminescence reaction. The most widely studied one, has advanced in several lineages of beetle's species and Coleoptera families including fireflies (*Lampyridae*). The beetle's species (*Elateridae*) and railroad worms (*Phengodidae*) to create a glowing emitting light with 540-640nm wavelength ranging from a green to red fluorescent light. These chemical reactions utilise a stable and non-toxic compound (Tiffen *et al.*, 2010, Viviani *et al.*, 2011, Kotlobay *et al.*, 2020).

Firefly luciferase can be expressed self-sufficiently or fused with proteins of interest in several plant organs, for example roots, stems, and leaves, during the advance transgenic plant development in the area of research (Li *et al.*, 2021). Affords a useful tool to study protein expression and localization, and protein-protein interaction (PPIs). Firefly luciferin catalyzing firefly luciferase gene in the occurrence of magnesium (Mg^{2+}), oxygen (O_2), and ATPs, and produce by the yellow-green luminescence. Coenzyme A (CoA) could be used to deoxidize the oxidized light emitting compound found in living organism that generate bioluminescence (Gosset *et al.*, 2020).

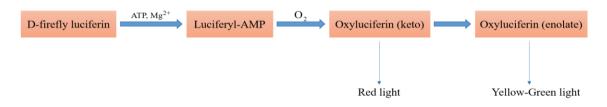


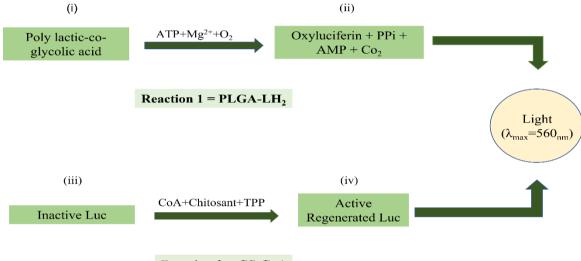
Figure 2: The firefly luciferase bioluminescent rection

II. Glowing plant in Pressurized bath infusion of nanoparticles (PBIN)

In 2017, a novel nano-technique called pressurized bath infusion of nanoparticles (PBIN) techniques. PBIN method works by supplying an external pressure against the internal microchannels with in the leaf mesophyll cells. In this technique, to introduce the firefly luciferin and Coenzyme A (CoA) into two separate nano capsules, and luciferase connected onto the outer layer of silica-PEG nanocarriers. These nanoparticles could decrease luciferin toxicity and artificial denaturation of luciferase, producing a long light-emitting degeneration ratio of up to 8 hours in plant cells.



In this work, to insert the nanoparticles mixture into the entire plant cells, the additional advance development a process of infusion using stomatal pores within the leaves. To develop a nano bionic plant with four separates nanoparticles types containing the protein and development necessary for self-sustaining photonic sources for direct and indirect light source. These different chemically interacting nanoparticles target the plant mesophyll cell and stomata guard cell, which is normally high for ATP. The preparation of nanoparticle system for producing light as per the following: (1) an immobilized silica nanoparticle (SNP-Luc) formed to a poly (ethylene glycol) bis (amine) (NH₂-PGE-NH₂, Mw 2,000, Sigma), (2) D-luciferin releasing poly (lactic-co-glycolic acid) (PLGA-LH₂) was synthesized the light-emitting compound luciferin, (3) Chitosan nanoparticles (medium Mw, Sigma) was added Coenzyme-A (CS-CoA) and CS-CoA was slowly added tripolyphosphate (TPP, Sigma). CoA is capable of release the light emission by regenerating firefly luciferase, reaction with dehydroluciferyl-adenylate, and (4) a silica nanoparticles. To introduce the nanoparticle mixture into the entire plant cell via a Pressurized Bath Infusion of Nanoparticles (PBIN) technique. Here, the whole plant is briefly merged in a pressured aqueous chamber. PBIN was successfully applied in saturation pressure of 1.8 bar without any membrane damage (Kwak *et al.*, 2017).



Reaction 2 = CS-CoA

Figure 3: Reaction mechanism of light production by firefly luciferase using Nanoparticles

III. Bacterial bioluminescence system

First bacterial bioluminescent system was reported in *Nicotiana tabacum*, only few bacterial luciferase-luciferin pairs were reported in other transgenic plants (Krichevsky *et al.*, 2010). All bioluminescence bacteria present the same type of mechanism for creating luminating brighter light, where photons are produced in a set of biochemical reactions utilizes flavin mononucleotide FMN and long chain aldehyde group derivative. Although bacterial bioluminescent genes can be target into plastids to engineer autoluminescence plant. This bacterial bioluminescence system is originated from the three gram-negative motile rods luminescent bacterium that is *Photobacterium, Vibrio and Xenorhabdus* (Piyush *et al.*, 2022). Luminous bacterial light-emission enzymatic catalyzing luciferin biosynthesis system is encoded by the *lux* operon. The highly conservative *lux* operons (*lux*CDABEG) are utilising the some unique and highly conserved among various species of all luminous bacteria, which encode the complete bacterial bioluminescent system (Close *et al.*, 2009). Gene *lux*A and *lux*B encode α and β subunits of the bacterial luciferase, and *lux*G encode for flavin reductase. Transfer the whole *Photobacterium leiognathi lux* operon into tobacco leaves chloroplast and fined the first auto-luminescent plant (Krichevsky *et al.*, 2010). Bacterial DNA gel blot analysis confirms integration of aadA and the *lux* operon in the *LUX*-rps12/TrnV. Total cellular DNA isolated from transplastomic leaves, the TrnA and TrnI plasmid geans aadA and selective spectinomycin resistance gene (Krichevsky *et al.*, 2010).

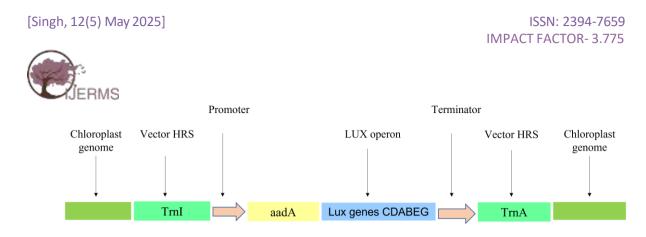


Figure 4: LUX-TrnI/TrnA transplastomic chloroplast genome

IV. Fungal bioluminescent system

Recently; the fourth bioluminescent system originated by bioluminescence fungi has been describe to produce strong selfsustain glowing plant, providing the first genetically encodable pathway from eukaryotes. The fungal bioluminescent system produces emitting light and shows no cytotoxicity and growth inhibition in plant cells. (Kotlobay *et al.*, 2018). It is a unique and genetically encodable eukaryotic luciferase-luciferin system, it was introduced into plant cell (Khakhar *et al.*, 2020; Mitiouchkina *et al.*, 2020). The biosynthetic pathway of fungal luciferin is called caffeic acid cycle (present in all plants) and report self-sustain bioluminescence that is visible to the naked eyes giving a clear lighting asset. Caffeic acid cycle is a metabolic pathway responsible for luminescence in fungi.

To develop transgenic tobacco plants (*Nicotiana benthamiana*) with advance features were performed continuously expressing fungal caffeic acid cycle and taken from the luminescent fungus *Neonothopanus nambi*. The process followed by four types of important enzymes that catalyse the caffeic acid cycle derivative pathways: fungal luciferase (Luz), hispidin synthase (HispS), and hispidin-3-hydroxylase (H3H), and caffeylpyruvate hydrolase (CPH). Another enzyme is NPGA (4'-phosphopantetheinyl transferase), initiates HispS post-translationally (Ke and Tsai 2022). Tree fungus Omphalotus olearius produce a bright glow at night (Farusi and Watt 2016). Glowing plant expressing emitting light for these genes during all developmental stages and the most important think is – no need for any type of external substrates to express enough brightness to be easily visible light. The self-luminescent tobacco plant was brighter than other reported self-light-emitting plants (Khakhar *et al.*, 2020).

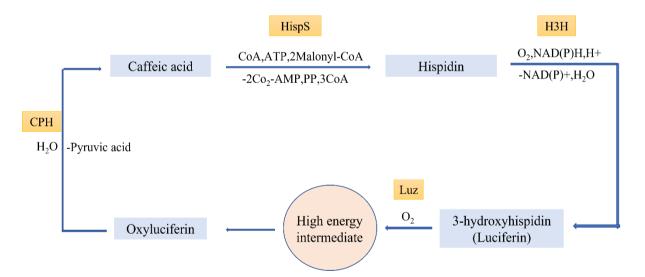


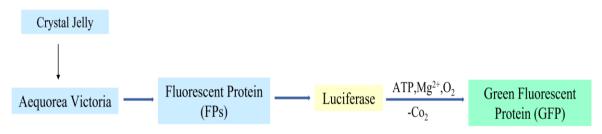
Figure 5: Fungal luciferin biosynthetic bioluminescence pathway

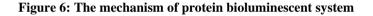
Working on caffeic acid cycle (CAC): The caffeic acid cycle is a native biosynthetic pathway in all plants, and does not appear to harm the plant. Caffeic acid cycle generally present in all plant cell. Fungal bioluminescence system, expression of Caffeic acid cycle is not toxic in plants. Mainly it is a metabolic process for the fungus luciferin. Fungal luciferin is converted into 3-hydroxyhispidin. Luz oxidises luciferin to produce photons and caffelpyruvic acid to hydrolyzed and produce caffeic acid and pyruvic acid. HispS (hispidin synthase) post-translationally can recycle caffeic acid to make hispidin by Hips (Purtov *et al.*, 2015 and Oba *et al.*, 2017). Three intermediary metabolites lignins, flavonoid anthocyanins condensed tannins, and shikimate produced by the fungal caffeic acid cycle (Mitiouchkina *et al.*, 2020 and Li *et al.*, 2021).



V. Protein bioluminescent system

In 1995, green fluorescent protein (GFP) gene isolate from jellyfish *Aequorea Victoria* was expressed in *Arabidopsis*, novel research in living glowing plant cell (Hu and Cheng 1995). Protein modification change into amino acid sequences of luciferase can lead to significant shifts in emission wavelength. The photoprotein present in them is called GFP. Fluorescent protein (FPs) is covering various type of fluorescent range (i.e., yellow, blue, green, red, and cyan,) have been modified for plant cells (Blatt and Grefen 2014). Another practically significant group of bioluminescent reactions. A few plants have recently been converted using bioluminescence systems such as the FPs are responsible for induction of most native luminescence (Kim and Paulmurugan 2021). FPs produce photoluminescence that allow them to glow without consumption of ATP (Chudakov *et al.*, 2010). Bioluminescent system glowing light by oxidation of a specific substrate, luciferin, with co-factors, i.e. ATP and NADH and catalyzed by the specific enzymes, luciferase (Fleiss and Sarkisyan 2019). Bioluminescent protein aequorin to excite GFP. The electrons of FPs are promoted into electronic excited state by a radiation less energy transfer and then relax with they emit light production (Rowe *et al.*, 2009).

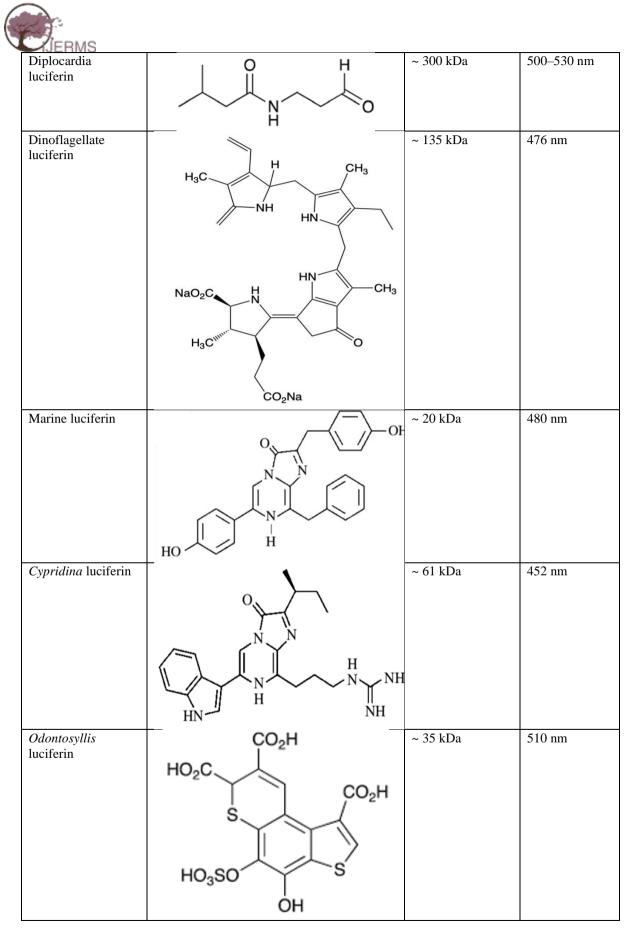




Protein bioluminescent system provide two representatives system – nano luciferase (NanoLuc) (England *et al.*, 2016) and nano lantern (Doerr, 2013). Nano luciferase isolate from shrimp *Oplophorous gracilirotris* luciferase (Hall *et al.*, 2012). Nano lantern is an artificial luminescent protein molecule fused with enhanced *Renilla reniformis* luciferase (Saito *et al.*, 2012). Recently, green enhance fluorescence nano-lantern gene was introduce into *A. thaliana*, Luminescence became outstanding brighter than suitable substrate furimazine (Furuhata *et al.*, 2020).

Bioluminescence	Luciferin	Luciferase Size	Wavelength
Species			_
Bacterial luciferin		~ 75 kDa	490 nm
Fungal luciferin	он	~ 29 kDa	520 nm
	ОН		
	но		
	ОН		
D - luciferin	HO	~ 280 kDa	560 nm

Table 2: Molecular structure of the known Luciferins gene





Diversity of Bioluminescent Systems: bioluminescence has independently evolved over 40 times in nature (Haddock *et al.*, 2010), resulting in varied chemical systems and colours of emitted light. The diversity and mechanisms of bioluminescence systems across various organisms:

Organism Type	Example Species	Emission Colour	System Type
Bacteria	Vibrio fischeri	Blue-Green	FMNH2-dependent
Fungi	Neonothopanus nambi	Green	ATP-independent
Insects	Photinus pyralis (firefly)	Green-Yellow	ATP-dependent luciferase
Marine organisms	Aequorea victoria	Blue	Photoproteins (i.e. aequorin)
Dinoflagellates	Noctiluca scintillans	Blue	Luciferin-binding
			proteins

Table 3: Variety of Bioluminescence Organisms and Mechanisms

Colours of Bioluminescence: In nature, bioluminescence produces different colours of light – mainly blue, green, red, yellow, orange, turquoise, and purple. Blue emission is most common bioluminescent colour, especially prevalent in marine organisms such as jellyfish, dinoflagellates, and deep-sea fish. Blue light penetrates water effectively, making it ideal for communication and predation in oceanic environments. Green emission found in some beetles and fungi. Green bioluminescence is often associated with terrestrial organisms, such as *Mycena chlorophos* and *Panellus stipticus*, or engineered systems expressing GFP (green fluorescent protein). Yellow to Orange emission observed in some fireflies and other insects. Variations in the luciferase structure among firefly species cause emission shifts from green to orange. Red emission light is extremely rare in nature. One notable example is the deep-sea dragonfish (*Malacosteus niger*), which can emit and detect red bioluminescence, giving it a stealth advantage in the deep ocean. Turquoise bioluminescence characterized by light emission in the 475-495nm wavelength range is a captivating phenomenon with significant applications in bioimaging optogenetics and synthetic biology. Purple bioluminescence emitting light in the 400-420nm range is relatively rare in nature. This emission is typically achieved through specific luciferase-luciferin system, engineered photoproteins or synthetic substrates.

Luciferin	Luminescence maximum (nm)	Approximate colour	
Coelenterazine	400 nm	Purple	
	450-480 nm	Blue to Turquoise	
Dinoflagellate luciferin	474 nm	Blue	
Bacterial luciferin	490 nm	Turquoise	
Fungus luciferin	520-530 nm	Yellow	
Firefly luciferin	560 nm	Green	
	615 nm	Orange	
Deep sea dragonfish	750 nm	Red	

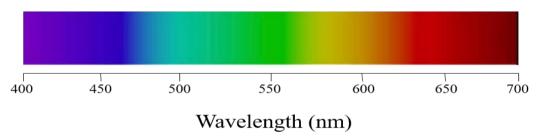
 Table 4: Emission Wavelengths of Bioluminescence in Various Organisms

In India, over 100 species across various taxa exhibit bioluminescence including fireflies (*Lampyridae*), click beetles (*Elateridae*), and certain fungi. These organisms utilize bioluminescence for purposes such as mating communication and predation. Despite their ecological significance studies on their distribution and applications in India are limited highlighting the need for further research in biodiversity and conservation (Chatragadda 2020). Certain fungi such as Armillaria mellea and Mycena chlorophose exhibit bioluminescence. The light emission is believed to attract insects aiding in spore dispersal (Duchatelet and Dupont 2024).



Organism	Common Name	Wavelength range of light	
0		emission	
Aequorea aequorea	Jelly fish	~ 500-523nm	
Aequorea victoria	Jelly fish	~ 520nm	
Diplocardia longa	Earthworm	~ 500-530nm	
Gonyulax polyedra	Dinoflagellate	~ 479nm	
Mycena mushroom	Fungal	~ 520-530nm	
Odontosyllis luciferin	Fireworm	~ 510nm	
Photinus pyralis	Firefly	~ 530-590nm	
Pleurotus japonicus	Moon night mushroom	~ 524nm	
Vibrio fischeri	Marine bacterium	~ 489nm	
Malacosteus niger	Deep sea dragonfish	~ 750nm	

The Visible Spectrum



Applications of Bioluminescence Systems

- 1. Biomedical Imaging => luciferase reporters are widely used in non-invasive in vivo imaging to track gene expression, cancer progression, and infection dynamic. Bioluminescence imaging (BLI) is a powerful technique used to monitor cellular and molecular processes in living animals. Luciferase-expressing cells or tissues can be tracked over time (Greer and Szalay 2002).
- 2. Biosensing => bioluminescent systems are applied in environmental monitoring for detecting toxins, heavy metals, and pollutants. Whole-cell biosensors expressing luciferase genes respond to specific analytes with a light signal. Engineered microorganisms expressing bioluminescent gene can detect and respond to environmental contaminants including - Heavy metals (i.e. mercury, cadmium), Organic pollutants (i.e. toluene, pesticides), and Antibiotics in water sources (Van der Meer and Belkin 2010).
- 3. Synthetic Biology => engineered systems using fungal or bacterial bioluminescence have been incorporated into plants, bacteria, and mammalian cells for real-time biological feedback and smart biosystems (Mitiouchkina et al., 2020).
- 4. **Sustainable Technologies** => bioluminescent systems are being explored for use in eco-friendly lighting especially in genetically engineered plants or algae as alternative to electricity-based lights in public spaces (Escher et al., 2020).
- 5. **Ecological Monitoring** => natural or synthetic bioluminescent organisms can be used to: Monitor ecosystem health and Track biogeochemical cycles in marine environments (Haddock et al., 2010).
- 6. Engineered bioluminescence Plants and Algae => recent innovations have enabled genetically engineered plants and algae to autonomously glow, potentially serving as: Living lights for sustainable urban design (Escher et al., 2020) and Biosensore for plant health or environmental stress (Mitiouchkina et al., 2020).

Use of Bioluminescence plant in society

The most important applications of this system are living bacteria, fungi, insects, firefly, are utilised as a light source. The main applications of the bioluminescence system are the studies of antimicrobial drugs, bacterial infections, and environmental monitoring (Bjorkman and Karl 2001). Fluorescent Protein can provide the fluorescent flowers and fluorescent plant specimens. Fluorescent plants utilized as alternative light source for desk lamp or street bulb in decorative area (Dunuweera et al., 2024). It is not only to produce a glowing street bulb, but the fluorescent plant will also play a major role in this solar energy. Transgenic plants are capable to produce auto luminescence light with new technology will allow researchers can study environmental stress in bioluminescence plant metabolism. Glowing plant can be supplied in the market, so that electricity will work for it and it will also be helpful in the economy. Indoor lighting; we can't live without it, on the one hand light is becoming very expensive and it is also bad for the environment.



Use as a sustainable light source in the future prospective

In future possibilities; production of transgenic bioluminescence plant can be an alternative option as decorative stuff for buildings and areas. Engineered bioluminescence plants through synthetic biology to autonomously emit light without external energy input, offer a promising, nature-based alternative for low-energy lighting. Also, create ornamental glowing plant as a street or reading lamp, which doesn't use any electrical energy. Glowing light will produce automatically by the metabolism activities of living plant leaf. The innovation will lead to low intensity indoor lighting and also transformation into these plants or trees into self-made light and all the streetlights and pole lights. Somehow this alternative source of lighting will also help to conserve lights on large or small scale. These glowing plants are very useful to promoting substitutes of artificial light to light up the night sky in the future.

In this article, we provide an intentionally brief overview of emitting light reaction. Thus, they have done with many plants like kale, spinach, arugula, and watercress can be used as the illumination for reading and writing.

Sustainable development in the next generation of society

In the future; sustainable development has evolved from a concept into imperative for global well-being. Defined by the Brundtland Commission as development that "meets the needs of the present without compromising the ability of future generations to meet their own needs" it emphasizes the balanced integration of economic growth, environmental protection, and social equity. Farmers can cultivate this plant, which will be a good source of income. These bioluminescence plants can also be used as ornamentals, which are largely for decoration purpose. This glowing plant will prove to be a good and beautiful result of the research area and will also provide an advantage field of research. These bioluminescence plants will be very attractive and will prove to be a good example of novel research, so they will also be expensive. If this is an ornamental plant – then its price will also be high, which will prove beneficial for the economy condition and farmers will also get benefit. This bioluminescent plant will prove to be helpful in the economy as will as it will save some amount of electricity, which is also good for our environment. As the next generation faces intensifying challenges – climate change, resource depletion, biodiversity loss, and social inequalities – sustainable development must transform from policy to practice across all sectors of society.

3. CONCLUSION

The concept of bioluminescence plants represents a visionary intersection of synthetic biology, environmental design, and sustainable innovation. Bioluminescence is a simple technique for generating emitted light that produces light in the presence of oxygen. Light can also be generated in living organisms by transferring the bioluminescence gene. Light is produced by transferring the genes of many types of bacteria, fungi, insects, firefly, marine organism, algae, dinoflagellate, jellyfish, earthworm, and sea star fish to the living organism. By integrating natural light-producing pathways into plant systems, researchers are exploring the possibility of creating living, self-sustaining sources of light that operate without the need for external electricity or chemical inputs. These genetically engineered organisms could significantly contribute to energy-efficient urban lighting, eco-friendly interior design and real-time environmental monitoring, aligning closely with the core principles of sustainable development. In the last several years many types of luminescence systems have been discovered such as – the luciferase-luciferin system, PBIN technique, bacterial bioluminescence system, caffeic acid cycle (CAC) and fluorescent proteins (FPs), all these techniques can convert normal plant into glowing plant which can be seen with naked eyes. In this review, attention the focuses will be the roll of bioluminescent system. In this review paper; a brief summary of almost all the techniques used to make a bioluminescence system will likely play an increasingly prominent role in next-generation bio-integrated technologies.

REFERENCES

- 1. Bjorkman K.M., and Karl D.M., (2001). Novel method for the measurement of dissolved adenosine and guanosine triphosphate in aquatic habitats: applications to marine microbial ecology. J Microbial Methods. 47(2):159-167.
- 2. Blatt M.R., and Grefen C., (2014). Arabidopsis Protocols. Totowa, NJ: Humana Press. 487-507.
- 3. Chatragadda R., (2020). Terrestrial and marine bioluminescence organisms from the Indian subcontinent: a review. Pubmed. 192(12):747. DOI:10.1007/s10661-020-08685-5.
- 4. Chin D.P., Shiratori I., Shimizu A., Kato K., Mii M., and Waga I., (2018). Generation of brilliant green fluorescent petunia plants by using a new and potent fluorescent protein transgene. Scientific Reports. 8(1):16556.
- 5. Chudakov D.M., Matz M.V., Lukyanov S., and Lukyanov K.A., (2010). Fluorescent Proteins and their applications in imaging living cells and tissues. Physiological reviews. 90(3):1103-1163. DOI:10.1152/Physrev.00038.2009.
- 6. Close D.M., Ripp S., and Sayler G.S., (2009). Reporter proteins in whole-cell optical bioreporter detection system, biosensor integration and biosensing applications. Sensors. 9(11):9147-9174.
- 7. Doerr A., (2013). Nano-lantern lights the way. Nature Methods. 10(2):104-104. DOI:10.1038/nmeth.2356.



- 8. Duchatelet L., and Dupont S., (2024). Marine eukaryote bioluminescence: a review of species and their functional biology. Marine Life Science & Technology. https://doi.org/10.1007/s42995-024-00250-0.
- Dunuweera A.N., Dunuweera S.P., and Ranganathan K., (2024). A Comprehensive Exploration of Bioluminescence System, Mechanisms, and Advanced Assays for Versatile Applications. Biochemistry Research International Volume 2024, Article ID 8273237, 22 pages. https://doi.org/10.1155/2024/8273237.
- 10. England C.G., Ehlerding E.B., and Cai W., (2016). NanoLuc: A small luciferase is brightening up the field of bioluminescence. Bioconjugate Chem. 27(5):1175-1187. DOI:10.1021/acs.bioconjchem.6b00112.
- 11. Escher A., Woudenberg D., and Roelfsema M.R.G., (2020). Bioluminescent plants: Lighting up urban landscapes with synthetic biology. Trends in Biotechnology, 38(7):721-734.
- 12. Farusi G., and Watt S., (2016). Living light: the chemistry of bioluminescence. School and Science in School. 35:30-36.
- 13. Fleiss A., and Sarkisyan K.S., (2019). A brief review of bioluminescent systems (2019). Current genetics. 65(4):877-882. DOI:10.1007/s00294-019-00951-5.
- 14. Furuhata Y., Sakai A., Murakami T., Nagasaki A., and Kato Y., (2020). Bioluminescent imaging of Arabidopsis thaliana using an enhanced Nano-lantern luminescence reporter system. PLOS One. 15(1):e0227477. DOI:10.1371/journal.pone.0227477.
- 15. Gordiichuk P., Coleman S., Zhang G., Kuehne M., Lew T.T.S., Park M., Cui J., Brooks A.M., Hudson K., Graziano A.M., Marshall D.J.M., Karsan Z., Kennedy S., and Strano M.S., (2021). Augmenting the living plant mesophyll into a photonic capacitor. Science Advance. 7:eabe9733.
- 16. Gosset P., Taupier G., Cregut O., Brazard J., Mely Y., Dorkenoo K., Leonard J., and Didier P., (2020). Excitedstate proton transfer in oxyluciferin and its analogues. The Journal of Physical Chemistry Letters. 11(9):3653-3659.
- 17. Greer L.F., and Szalay A.A., (2002). Imaging of light emission from the expression of luciferases in living cells and organisms: A review. Luminescence 17(1):43-74.
- 18. Haddock S.H., Moline M.A., and Case J.F., (2010). Bioluminescence in the sea. Annual Review of Marine Science. 2(1):443-493.
- Hall M.P., Unch J., Binkowski B.F., Valley M.P., Butler B.L., Wood M.G., Otto P., Zimmerman K., Vidugiris G., Machleidt T., Robers M.B., Benink H.A., Eggers C.T., Slater M.R., Meisenheimer P.L., Klaubert D.H., Fan F., Encell L.P., and Wood K.V., (2012). Engineered luciferase reporter from a deep-sea shrimp utilizing a novel imidazopyrazinone substrate. ACS chemical biology. 7(11):1848-1857. DOI:10.1021/cb3002478.
- 20. Hu W., and Cheng C., (1995). Expression of Aequorea green fluorescent protein in plant cells. FEBS Latters. 369:331-334. DOI:10.1016/0014-5793(95)00776-6.
- 21. John L. W., (2008). Bioluminescence: the first 3000 years (Review). Journal of Siberian Federal University. Biology 3(1):194-205.
- 22. Karunarathna S.C., Mortimer P.E., Tibpromma S., Dutta A.K., Paloi S., Hu Y., Baurah G., Axford S., Marciniak C., Luangharn T., Madawala S., Lin C., Chen J., Acharya K., and Kobmoo N., (2020). Roridomyces phyllostachydis (Agaricales, Mycenaceae), a new bioluminescent fungus from Northeast India Phytotaxa. 459:155-167. DOI:10.11646/Phytotaxa.459.2.6.
- 23. Ke H.M., and Tsai I.J., (2022). Understanding and using fungal bioluminescence Recent progress and future perspectives. 33:100570. DOI:10.1016.
- 24. Khakhar A., Starker C.G., Chamness J.C., Lee N., Stokke S., Wang C., Swanson R., Rizvi F., Imaizumi T., and Voytas D.F., (2020). Building customizable auto-luminescence luciferase-based reporters in plants. eLife. 9:e52786.
- 25. Kim S.B., and Paulmurugan R. (2021). Bioluminescent imaging systems for assay developments. Analytical Sciences 37(2):233-247. DOI:10.2116/Analsci.20R003.
- 26. Kishi-Kaboshi M., Aida R., and Sasaki K., (2017). Generation of gene-edited chrysanthemum morifolium using multicopy transgenes as targets and markers. Plant and Cell Physiology. 58(2):216-226.
- 27. Kotlobay A.A., Kaskova Z.M., and Yampolsky I.V., (2020). Palette of luciferases: natural for new applications in biomedicine. Acta Nat. 12:15-27. DOI:10.322607/actanaturae.10967.
- 28. Kotlobay A.A., Sarkisyan K.S., Mokrushina Y.A., Marcet-Houben M., Serebrovskaya E.O., Markina N.M., Somermeyer L.G., Gorokhovatsky A.Y., Vvedensky A., Purtov K.V., Petushkov V.N., Rodionova N.S., Chepurnyh T.V., Fakhranurova L.I., Guglya E.B., Ziganshin R., Tsarkova A.S., Kaskova Z.M., Shender V., Abakumov M., Abakumova T.O., Povolotskaya I.S., Eroshkin F.M., Zaraisky A.G., Mishin A.S., Dolgov S.V., Mitiouchkina T.Y., Kopantzev E.P., Waldenmaier H.E., Oliveira A.G., Oba Y., Barsova E., Bogdanova E.A., Gabaldon T., Stevani C.V., Lukyanov S., Smirnov I.V., Gitelson J.I., Kondrashov F.A., and Yampolsky I.V., (2018). Genetically encodable bioluminescent system from fungi. Proceedings of the National Academy of Sciences. 115(50):12728-12732. DOI:10.1073/pnas.1803615115.

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- 29. Krichevsky A., Meyers B., Vainstein A., Maliga P., and Citovsky V., (2010). Autoluminescence plants. Ed. Vladimie N. uversky. PLOS One. 5(11):e15461. DOI:10.1371/journal.pone.0015461.
- 30. Kwak S., Giraldo J.P., Wong M.H., Koman V.B., Lew T.T.S., Ell J., Weidman M.C., Sinclair R.M., Landry M.P., Tisdale W.A., and Strano M.S., (2017). A nanobionic light-emitting plant. Nano Letters. 17(12):7951-7961.
- 31. Li B., Chen R., Zhu C., and Kong F., (2021). Glowing plants can light up the night sky? A review Biotechnology and Bioengineering. 118(10):3706-3715.
- 32. Mitiouchkina T., Mishin A.S., Somermeyer L.G., Markina N.M., Chepurnyh T.V., Guglya E.B., Karataeva T.A., Palkina K.A., Shakhova E.S., Fakhranurova L.I., Chekova S.V., Tsarkova A.S., Golubev Y.V., Negrebetsky V.V., Dolgushin S.A., Bubyrev A.I., Pushin A.S., Choob V.V., Dolgov S.V., Kondrashov F.A., Yampolsky I.V., and Sarkisyan K.S., (2020). Plants with genetically encoded auto luminescence. Nature Biotechnology. 809376.
- 33. Morise H., Shimomura O., Johnson F. H., and Winant J., (1974). Intermolecular energy transfer in the bioluminescent system of Aequorea. Biochemistry 13:2656-2662.
- 34. Oba Y., Suzuki Y., Martins G.N.R., Carvalho R.P., Pereira T.A., Waldenmaier H.E., Kanie S., Naito M., Oliveira A.G., Dorr F.A., Pinto E., Yampolsky I.V., and Stevani C.V., (2017). Identification of hispidin as a bioluminescent active compound and its recycling biosynthesis in the luminous fungal fruiting body. Photochemical and Photobiological Sciences. 16(9):1435-1440. DOI:10.1039/C7PP00216E.
- 35. Ow D. W., DE Wet J. R., Helinski D. R., Howell S. H., Wood K. V., and Deluca M., (1986). Transient and stable expression of the firefly luciferase gene in plant cells and transgenic plants. Science. 234(4778):856-859.
- 36. Piyush., Verma S., Gautam A., and Rani N., (2022). A brief review on glowing plants and involvement of different bioluminescence systems. International Journal of Botany Studies. 2455-541X:25-33.
- Purtov K.V., Petushkov V.N., Baranov M.S., Mineev K.S., Rodionova N.S., Kaskova Z.M., Tsarkova A.S., Petunin A.I., Bondar V.S., Rodicheva E.K., Medvedeva S.E., Oba Y., Arseniev A.S., Lukyanov S., Gitelson J.I., and Yampolsky I.V., (2015). The chemical basis of fungal bioluminescence. Angewandte Chemie International Edition. 54(28):8124-8128. DOI:10.1002/anie.201501779.
- 38. Rowe L., Dikici E., and Daunert S., (2009). Engineering Bioluminescent Proteins: Expanding their Analytical Potential. Anal Chem. 1;81(21):8662-8668. DOI:10.1021/ac9007286.
- 39. Saito K., Chang Y., Horikawa K., Hatsugai N., Higuchi Y., Hashida M., Yoshida Y., Matsuda T., Arai Y., and Nagai T., (2012). Luminescent proteins for high-speed single-cell and whole-body imaging. Nature Communications. 3(1):1262.
- 40. Sasaki K., Kato K., Mishima H., Furuichi M., Waga I., Takane K., Yamaguchi H., and Ohtsubo N., (2014). Generation of fluorescent flower exhibiting strong fluorescence by combination of fluorescent protein from marine plankton and recent genetic tools in Torenia fournieri Lind. Plant Biotechnology. 31(4):309-318.
- 41. Shimomura O., (2006). Bioluminescence: Chemical Principles and Methods. DOI:10.1142/6102. ISBN: 978-981-256-801-4.
- 42. Tiffen J.C., Bailey C.G., Ng C. Rasko J.E.J., Holst J., (2010). Luciferase expression and bioluminescence does not affect tumor cell growth in vitro or in vivo. Mol Cancer 9:299.
- 43. Van der Meer J.R., and Belkin S., (2010). Where microbiology meets microengineering: Design and applications of reporter bacteria. Nature Reviews Microbiology 8:511-522.
- 44. Viviani V.R., Amaral D., Prado R., and Arnoldi F.G.C., (2011). A new blue-shifted luciferase from the Brazilian *Amydetes fanestratus* (Coleoptera:Lampyridae) firefly: molecular evolution and structural/functional properties. *Photochem. Photobiol.* Sci. 10:1879-1886. DOI:10.1039/clpp05210a.